Exposure to fine particulate matter and semen quality in Taiwan

Xiang Qian Lao,1,2 Zilong Zhang,1 Alexis K H Lau,3,4 Ta-Chien Chan,5 Yuan Chieh Chuang,6 Jimmy Chan,3 Changqing Lin,4 Cui Guo,1 Wun Kai Jiang,6 Tony Tam,7 Gerard Hoek,8 Haidong Kan,9 Eng-kiong Yeoh,1 Ly-yun Chang6,10

ABSTRACT

Objectives Environmental exposure to chemicals has been considered a potential factor contributing to deteriorated semen quality. However, previous literature on exposure to air pollution and semen quality is inconsistent. We therefore investigated the health effects of short-term and long-term exposure to fine particulate matter (PM1.3) on semen quality in Taiwanese men from the general population.

Methods A cross-sectional study was conducted among 6475 male participants aged 15 to 49 years who participated in a standard medical examination programme in Taiwan between 2001 and 2014. Semen quality was assessed according to the WHO 1999 guideline, including sperm concentration, total motility, progressive motility and morphology. Three-month and 2-year average PM1.3 concentrations were estimated at each participant’s address using a spatiotemporal model based on satellite-derived aerosol optical depth data. Multivariable linear and logistic regressions were used to examine the associations between PM1.3 and semen quality.

Results A robust association was observed between exposure to PM1.3 and decreased normal morphology. Every increment of 5 µg/m3 in 2-year average PM1.3 was significantly associated with a decrease of 1.29% in sperm normal morphology and a 26% increased risk of having the bottom 10% of sperm normal morphology, after adjusting for a wide range of potential confounders (p<0.001). On the other hand, an increment of 5 µg/m3 in 2-year average PM1.3 was associated with an increase of 1.03×10^6/mL in sperm concentration and a 10% decreased risk of being the bottom 10% of sperm concentration (both p<0.001). Similar results were found for 3-month PM1.3.

Conclusions Exposure to ambient PM1.3 air pollution is associated with a lower level of sperm normal morphology and a higher level of sperm concentration.

INTRODUCTION

Infertility is a global public health concern. It was estimated that 48.5 million couples worldwide were infertile in 2010,2 with men contributing >50%.3 Environmental exposure to chemicals, including air pollutants, has been considered as one of the potential factors contributing to deteriorated semen quality,4 although the biological mechanisms remain uncertain. Many of the components of ambient particulate matter (PM), such as heavy metals and polycyclic aromatic hydrocarbons (PAHs), have been shown to have toxic effects on semen quality in animal experiments.4–6 Oxidative stress induced by particles is also of the potential to disturb spermatogenesis.

Air pollution has become the world’s largest single environmental health risk. Exposure to ambient air pollution was estimated to cause >3.7 million premature deaths worldwide in 2012.7 PM is the most important air pollutant and affects more people than any other pollutants. There has been a large volume of epidemiological studies examining air pollution and its various health outcomes including birth defects. However, only a few studies with small samples have investigated the health effects of ambient PM on semen quality in humans, and the results have been inconsistent.8–15 In addition, most of these studies focused on the acute or short-term effects, despite the fact that people generally suffer from prolonged exposure. Chronic, low-dose exposure may contribute to significant spermatogenesis impairment.16 Moreover, previous studies have generally estimated PM exposure using residential proximity to routine ground-level air pollution monitoring stations. The exposure is typically at the community (district, county or city) rather than the individual level, with the same exposure level assigned to all of a community’s. This limitation may mask the variation, or
cause misclassification, thus leading to the inconsistent results of possible effects associated with PM air pollution. We therefore investigated the health effects of exposure (including long-term exposure and a spermatogenic cycle exposure) to fine PM—PM with an aerodynamic diameter of <2.5 μm (PM$_{2.5}$)—on semen quality in Taiwanese men from the general population, using a spatiotemporal model based on satellite-derived aerosol optical depth (AOD) data to estimate each participant’s PM$_{2.5}$ exposure.

METHODS

Study population

We studied male participants from a large cohort consisting of Taiwanese residents who participated in a standard medical examination programme run by a private firm (MJ Health Management Institution, Taipei, Taiwan). The details have been documented elsewhere. $^{17}$ All procedures of the medical examination programme are approved according to ISO9001. Information on the medical examinations and related quality control can be accessed in the Technical Report of the MJ Health Research Foundation. $^{20}$ In brief, the participants received a series of medical examinations including general physical examinations, anthropometric measurements and biochemical tests of blood and urine. They also completed a standard self-administered questionnaire survey. The semen quality assessment was voluntarily selected by the male participants. All of the participants had similar age and semen parameters (all p>0.05). There were 158,542 male participants of reproductive age (15–49 years) $^{21}$ who participated in the programme from 2001 to 2014, when satellite AOD data from the US National Aeronautics and Space Administration were available for air pollution exposure estimation. During the study period, 6938 participants voluntarily selected a semen quality examination. Generally, participants receiving semen measurements were comparable with all the 158,542 male participants in terms of age, educational level, smoking habits and body mass index (BMI) (see online supplementary table 1). Only 211 participants (3.0%) had multiple visits with two or more semen tests, therefore, repeated-measures analysis was not conducted. A total of 463 participants with incomplete information were excluded (1 on anthropometric measures, 3 on blood test results, 174 on educational level, 236 on lifestyle factors and 49 on PM$_{2.5}$ exposure estimates due to missing address) and 6475 participants were ultimately included in data analysis. The 463 excluded participants had similar age and semen parameters (all p>0.05).

Semen analysis and health examination

All of the participants were required to be sexually abstinent 2–7 days before contributing semen samples. They were then instructed to collect semen samples in sterile containers by masturbation. If the sample was collected at home, the participant was required to mark down the time and send the sample to laboratory immediately. No assessments were conducted if the duration was longer than 2 hours after ejaculation. The semen samples were analysed at the MJ Health Screening Center’s central laboratory using standard protocols based on the WHO guidelines. $^{21}$ Sperm concentration, total motility (percentage of motile sperm), progressive motility (percentage of sperm with progressive motility) and morphology (percentage of sperm of normal morphology) were assessed by trained technicians. The assessment began as soon as the ejaculates had liquefied, and were completed within 1 hour of sample collection. An aliquot of the sample was put into a diluent solution and thoroughly mixed for the concentration assessment. A microcell counting chamber was used in six areas at a total microscope magnification of ×400. For the motility assessment, a 10 μL well-mixed sample from another aliquot was placed on a glass slide covered with a coverslip, and then examined at a total microscope magnification of ×400. Two hundred sperm cells were examined and classified into WHO motility categories A, B, C or D. The percentages of total motile sperm (A+B+C) and sperm with progressive motility (A+B) were calculated. Other smears were also prepared and air-dried, and then used for the morphology assessment. The WHO ‘strict’ criteria were used to examine 200 sperm cells and the percentage of morphologically normal sperm was calculated.

Details of health examination for other variables were described in previous publications. $^{17}$–$^{19}$

Ambient PM$_{2.5}$ estimate

A spatiotemporal model with high resolution (1×1 km) was developed based on satellite AOD data to retrieve ground-level PM$_{2.5}$ concentrations. The details have been described elsewhere. $^{17}$ This model was validated with ground-measured data from >70 monitoring stations in Taiwan from 2005 to 2014. (PM$_{1.0}$ data were only available for three monitoring stations from 2000 to 2004. Thus, validations were not conducted for this period.) The correlation coefficients ranged from 0.72 to 0.83 for yearly average PM$_{2.5}$ and 0.70 to 0.81 for monthly average PM$_{2.5}$ in different years (see online supplementary figure 1).

Each participant’s address was geo-coded into latitude and longitude, and address-specific monthly and yearly average PM$_{2.5}$ concentrations were calculated afterwards. The estimated annual average concentrations in Taiwan and at the participants’ locations are presented in figure 1. The southwestern and middle-eastern areas were generally the most and least heavily polluted, respectively. Most of the participants lived in the western area with apparent gradients of exposure. Because a spermatogenic cycle is around 3 months, we calculated 3-month average concentration (based on the

Figure 1 Map of the annual average concentration of satellite-based estimated PM$_{1.0}$ in Taiwan between 2001 and 2014. Circles represent the locations of the study participants. PM$_{1.0}$ particulate matter with an aerodynamic diameter <2.5 μm.
concentrations of the month of the medical examination and the 2 months before the medical examination) to assess the short-term effects. We also calculated 2-year average concentration (based on the concentrations of the year of the medical examination and the year before the medical examination) as an indicator of long-term exposure to ambient PM$_{2.5}$ air pollution.

Statistical analysis

We used multivariable linear regression models to evaluate the associations between PM$_{2.5}$ exposure and each semen quality parameter. Sperm concentration was log-transformed for data analysis due to its skewed distribution. A crude model without adjustment and three adjusted models were developed with potential confounders added gradually; model 1 was adjusted for age (years), education (high school or lower, college or university and postgraduate), smoking status (never, former and current), alcohol consumption (less than once/week, 1–3 times/week and >3 times/week), exercise (<1 hour/week, 1–2 hours/week and >2 hours/week) and occupational exposure to dust and organic solvents (yes and no); model 2 was further adjusted for BMI (calculated as weight (kg) divided by the square of height (m)), systolic blood pressure, fasting plasma glucose and total cholesterol (all as continuous variables); model 3 was further adjusted for season (calendar season) and year of the medical examination (categorical variable). Short-term and long-term effects of PM$_{2.5}$ exposure were analysed separately for comparison. The effect estimates were calculated for an increment of 5 µg/m$^3$ in average PM$_{2.5}$ concentrations. The potential effect modifications by age group (<35 years vs ≥35 years), smoking (never vs ever) and overweight (BMI ≥25.0 vs <25.0 kg/m$^2$) were examined by adding multiplicative interactions terms between PM$_{2.5}$ and these factors. Each potential modifier was examined in separate models.

We also used logistic regression models to investigate the associations between PM$_{2.5}$ exposure and the risk of decreased semen parameters. Participants with the bottom 10% of the parameters were defined as having decreased semen parameters. ORs were calculated for every 5 µg/m$^3$ increment in PM$_{2.5}$ concentrations.

We further performed subgroup analyses stratified by normal and abnormal semen parameters according to reference values from the WHO guidelines. A total of 1524 participants were categorised into abnormal group because they had at least one of the four semen parameters lower than the following reference limits: 15×10⁶/mL for sperm concentration, 40% for total motility, 32% for progressive motility and 4% for normal morphology. Consequently, the other 4951 participants were categorised into normal group.

All of the statistical analyses were performed using statistical package R V3.2.3 (R Core Team, Vienna, Austria). A two-tailed p value of <0.05 was considered to be statistically significant.

RESULTS

The general characteristics of the 6475 participants are presented in table 1. The average age was 31.9 years (SD: 4.3) and they were well educated. The majority never smoked (64.2%) and had alcohol use less than once a week (83.1%).

The semen characteristics by year are shown in table 2. For all of the participants, the mean (SD) values for sperm concentration, total motility, progressive motility and normal morphology were 54.0 (41.3)×10⁶/mL, 65.3 (14.5)%, 46.8 (16.1)% and 67.4 (14.9)%, respectively. Linear regression models were used to investigate the trends in semen parameters over the study period adjusting for age. An increasing trend was observed for progressive motility (p<0.001) and no trends were found for other three parameters (all p>0.05).

PM$_{2.5}$ distribution among the participants is shown by year in figure 2 and locations of the study participants are shown in figure 1. The PM$_{2.5}$ concentrations varied spatially among the participants within each year. The overall mean 3-month and 2-year PM$_{2.5}$ concentrations were 25.8 (SD: 8.5) and 26.1 (SD: 7.3) µg/m$^3$, with IQRs of 7.9 and 5.9 µg/m$^3$, respectively. The 3-month and the 2-year PM$_{2.5}$ concentration were highly correlated (correlation coefficient=0.81, p<0.001).

The results of the regression models are summarised in table 3. Both short-term and long-term exposure had similar effects on sperm concentration and morphology. An increment of 5 µg/m$^3$ in PM$_{2.5}$ was associated with a decrease in normal morphology (OR was 0.90 for both short-term and long-term exposure). In contrast, an increment of 5 µg/m$^3$ in PM$_{2.5}$ was associated with a decrease in normal morphology (0.83% for short-term exposure and 1.29% for long-term exposure) and increased risk of decreased normal morphology (ORs were 1.18 for short-term exposure and 1.26 for long-term exposure). There were no associations observed for progressive motility. Regarding the total motility, the association was not significant for short-term exposure in logistic regression analysis.

No significant effect modifications were observed for age group, smoking or overweight among the participants in relation to PM$_{2.5}$ concentration. Of the other factors, the potential effect modifications were observed by age group (ORs were 1.18 for short-term and 1.26 for long-term exposure) and increased risk of decreased normal morphology (0.83% for short-term exposure and 1.29% for long-term exposure) and reduced risk of decreased concentration (OR was 0.90 for both short-term and long-term exposure). In contrast, an increment of 5 µg/m$^3$ in PM$_{2.5}$ was associated with a decrease in normal morphology (0.83% for short-term exposure and 1.29% for long-term exposure) and increased risk of decreased normal morphology (ORs were 1.18 for short-term exposure and 1.26 for long-term exposure). There were no associations observed for progressive motility. Regarding the total motility, the association was not significant for short-term exposure in logistic regression analysis.
to the association between 3-month or 2-year PM$_{2.5}$ and semen parameters (all $p > 0.10$).

The subgroup analyses generally yielded similar results (see online supplementary table 2 for participants with normal semen parameters and online supplementary table 3 for participants with abnormal semen parameters).

**DISCUSSION**

To our knowledge, this is the largest study investigating the health effects of PM$_{2.5}$ air pollution on semen quality. It is also the first study considering the effects of both long-term exposure and short-term exposure (in a spermatogenic cycle). We find it is highly correlated between long-term and short-term exposure in this population. Because air pollution is quite stable over the study period, we speculate the exposure in a spermatogenic cycle is similar to the long-term exposure, and the effects are more likely the reflections of long-term exposure. We find robust associations between PM$_{2.5}$ exposure and semen quality. Every increment of 5 µg/m$^3$ in 2-year average PM$_{2.5}$ is associated with a decrease of 1.29% in sperm normal morphology, and a 26% increased risk of having the bottom 10% of sperm normal morphology, after adjusting for a wide range of potential confounders. In contrast, every increment of 5 µg/m$^3$ in 2-year average PM$_{2.5}$ is associated with an increase of 1.03×10$^6$/mL in sperm concentration and a 10% decreased risk of having the bottom 10% of sperm concentration. The associations remain robust in the analyses by taking into account a wide range of potential confounders, using different statistical methods (linear and logistic regression) and categorising participants into normal and abnormal groups.

The negative association between air pollution and normal morphology was also reported in the Teplice Program in Czech Republic, and in two recent studies in China and Poland. The average time course in these studies was 3-month concentration. However, long-term exposure was not taken into account. It is not clear if it was a short-term exposure or a surrogate of long-term exposure. Three studies in the USA, which the average time ranged from daily to 3-month concentration, did not observe significant associations. Hansen et al attributed the non-association to the relatively low level of air pollution.

The mechanism of how air pollution adversely affects sperm morphology is not fully understood. Air pollution-induced oxidative stress has been hypothesised to be one of the contributors to sperm dysfunction. A wide range of air pollutants, such as O$_3$, NO$_x$ and particles, can induce increased oxidative stress, which is related to the excessive production of reactive oxygen species (ROS). ROS and their metabolites can damage DNA, lipids and proteins, altering enzymatic systems and cell apoptosis and, ultimately, lead to decreasing semen quality. In addition, PM$_{2.5}$ may carry multiple trace elements and PAHs. The toxic effects of metals such as lead and cadmium on spermatogenesis have been well documented in animal models and confirmed in human.

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**Table 2** Distribution of semen parameters in Taiwanese men

<table>
<thead>
<tr>
<th>Year</th>
<th>Participants</th>
<th>Age (year)</th>
<th>Concentration ($10^6$/mL)*</th>
<th>Total motility (%)</th>
<th>Progressive motility (%)</th>
<th>Normal morphology (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2001</td>
<td>501</td>
<td>30.8 (4.2)</td>
<td>41.9 (2.3)</td>
<td>66.4 (13.7)</td>
<td>46.9 (15.3)</td>
<td>71.8 (13.7)</td>
</tr>
<tr>
<td>2002</td>
<td>611</td>
<td>31.1 (4.5)</td>
<td>41.8 (2.2)</td>
<td>65.5 (12.9)</td>
<td>45.7 (15.2)</td>
<td>66.5 (16.0)</td>
</tr>
<tr>
<td>2003</td>
<td>558</td>
<td>31.5 (4.3)</td>
<td>42.5 (2.3)</td>
<td>64.8 (14.3)</td>
<td>46.4 (15.6)</td>
<td>67.3 (16.1)</td>
</tr>
<tr>
<td>2004</td>
<td>535</td>
<td>31.1 (4.4)</td>
<td>40.6 (2.5)</td>
<td>63.3 (14.0)</td>
<td>48.4 (15.4)</td>
<td>67.9 (15.2)</td>
</tr>
<tr>
<td>2005</td>
<td>519</td>
<td>31.5 (3.9)</td>
<td>38.3 (2.4)</td>
<td>65.6 (14.2)</td>
<td>46.5 (15.4)</td>
<td>70.7 (13.5)</td>
</tr>
<tr>
<td>2006</td>
<td>530</td>
<td>31.3 (4.0)</td>
<td>39.1 (2.2)</td>
<td>64.8 (13.6)</td>
<td>45.7 (15.4)</td>
<td>67.5 (13.4)</td>
</tr>
<tr>
<td>2007</td>
<td>517</td>
<td>32.0 (3.8)</td>
<td>38.6 (2.6)</td>
<td>66.8 (12.7)</td>
<td>46.5 (15.1)</td>
<td>65.8 (13.5)</td>
</tr>
<tr>
<td>2008</td>
<td>556</td>
<td>32.3 (4.2)</td>
<td>38.6 (2.4)</td>
<td>64.8 (14.9)</td>
<td>45.9 (15.9)</td>
<td>65.8 (14.4)</td>
</tr>
<tr>
<td>2009</td>
<td>437</td>
<td>32.5 (4.4)</td>
<td>39.3 (2.5)</td>
<td>64.5 (14.8)</td>
<td>43.9 (14.4)</td>
<td>64.0 (14.3)</td>
</tr>
<tr>
<td>2010</td>
<td>447</td>
<td>32.7 (4.0)</td>
<td>39.8 (2.5)</td>
<td>64.2 (14.5)</td>
<td>44.4 (15.6)</td>
<td>63.7 (14.3)</td>
</tr>
<tr>
<td>2011</td>
<td>424</td>
<td>32.8 (4.2)</td>
<td>40.3 (2.3)</td>
<td>65.8 (15.4)</td>
<td>49.1 (18.0)</td>
<td>66.3 (14.9)</td>
</tr>
<tr>
<td>2012</td>
<td>402</td>
<td>33.0 (4.5)</td>
<td>44.3 (2.3)</td>
<td>64.2 (16.5)</td>
<td>48.1 (17.6)</td>
<td>67.8 (16.4)</td>
</tr>
<tr>
<td>2013</td>
<td>322</td>
<td>33.2 (4.1)</td>
<td>44.1 (2.2)</td>
<td>66.1 (16.6)</td>
<td>53.4 (19.7)</td>
<td>72.1 (13.1)</td>
</tr>
<tr>
<td>2014</td>
<td>116</td>
<td>32.7 (4.4)</td>
<td>45.9 (2.1)</td>
<td>64.1 (19.5)</td>
<td>47.1 (17.0)</td>
<td>77.0 (12.4)</td>
</tr>
<tr>
<td><strong>All</strong></td>
<td><strong>6475</strong></td>
<td><strong>31.9 (4.3)</strong></td>
<td><strong>40.6 (2.4)</strong></td>
<td><strong>65.3 (14.5)</strong></td>
<td><strong>46.8 (16.1)</strong></td>
<td><strong>67.4 (14.9)</strong></td>
</tr>
</tbody>
</table>

Results are presented as the mean (SD) for the age and semen parameters.

*p for trend

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**Figure 2** Distribution of PM$_{2.5}$ from 2001 to 2014; 3-month refers to the month of and the 2 months before the visit; 2-year refers to the year of and the year before the visit. Boxes cover the 25–75th percentile (IQR) with a centre line for the median concentration. Whiskers extend to the highest observation within 3 IQR of the box, with more extreme observations shown as circles. PM$_{2.5}$ particulate matter with an aerodynamic diameter <2.5 µm.

Table 3  Associations between exposure to PM2.5 and semen quality

<table>
<thead>
<tr>
<th>Semen parameters (n=6475)</th>
<th>Crude model*</th>
<th>Model 1*</th>
<th>Model 2*</th>
<th>Model 3*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Coef/OR (95% CI)† p Value</td>
<td>Coef/OR (95% CI)† p Value</td>
<td>Coef/OR (95% CI)† p Value</td>
<td>Coef/OR (95% CI)† p Value</td>
</tr>
<tr>
<td><strong>Short-term exposure (3-month average PM~2.5)</strong>*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Continuous</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Concentration (10^6 /mL)‡</td>
<td>1.04 (1.03 to 1.05) &lt;0.001</td>
<td>1.04 (1.02 to 1.05) &lt;0.001</td>
<td>1.04 (1.02 to 1.05) &lt;0.001</td>
<td>1.03 (1.02 to 1.05) &lt;0.001</td>
</tr>
<tr>
<td>Total motility (%)</td>
<td>0.28 (0.07 to 0.49) 0.008</td>
<td>0.18 (–0.02 to 0.39) 0.08</td>
<td>0.18 (–0.02 to 0.39) 0.08</td>
<td>0.19 (–0.02 to 0.41) 0.07</td>
</tr>
<tr>
<td>Progressive motility (%)</td>
<td>−0.02 (–0.25 to 0.21) 0.86</td>
<td>−0.08 (–0.31 to 0.15) 0.49</td>
<td>−0.07 (–0.30 to 0.16) 0.55</td>
<td>−0.05 (–0.29 to 0.19) 0.68</td>
</tr>
<tr>
<td>Normal morphology (%)</td>
<td>0.70 (-0.91 to –0.49) &lt;0.001</td>
<td>−0.78 (–1.00 to −0.57) &lt;0.001</td>
<td>−0.79 (–1.00 to −0.58) &lt;0.001</td>
<td>−0.91 (–1.13 to −0.69) &lt;0.001</td>
</tr>
<tr>
<td><strong>Dichotomous (&lt;10th percentile as decreased)</strong>*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Decreased concentration</td>
<td>0.88 (0.83 to 0.93) &lt;0.001</td>
<td>0.88 (0.83 to 0.93) &lt;0.001</td>
<td>0.88 (0.83 to 0.93) &lt;0.001</td>
<td>0.90 (0.85 to 0.95) &lt;0.001</td>
</tr>
<tr>
<td>Decreased total motility</td>
<td>0.94 (0.89 to 0.99) 0.02</td>
<td>0.95 (0.90 to 1.01) 0.10</td>
<td>0.95 (0.90 to 1.01) 0.10</td>
<td>0.97 (0.91 to 1.02) 0.27</td>
</tr>
<tr>
<td>Decreased progressive motility</td>
<td>0.96 (0.92 to 1.01) 0.16</td>
<td>0.98 (0.93 to 1.03) 0.42</td>
<td>0.98 (0.93 to 1.03) 0.44</td>
<td>1.00 (0.95 to 1.05) 0.94</td>
</tr>
<tr>
<td>Decreased normal morphology</td>
<td>1.16 (1.11 to 1.21) &lt;0.001</td>
<td>1.17 (1.12 to 1.22) &lt;0.001</td>
<td>1.17 (1.12 to 1.22) &lt;0.001</td>
<td>1.18 (1.13 to 1.24) &lt;0.001</td>
</tr>
<tr>
<td><strong>Long-term exposure (2-year average PM~2.5)</strong>*</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td><strong>Continuous</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Concentration (10^6 /mL)‡</td>
<td>1.03 (1.02 to 1.05) &lt;0.001</td>
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<td>1.03 (1.02 to 1.05) &lt;0.001</td>
</tr>
<tr>
<td>Total motility (%)</td>
<td>0.45 (0.21 to 0.69) &lt;0.001</td>
<td>0.32 (0.09 to 0.57) 0.008</td>
<td>0.33 (0.09 to 0.57) 0.007</td>
<td>0.34 (0.10 to 0.581) 0.006</td>
</tr>
<tr>
<td>Progressive motility (%)</td>
<td>0.13 (–0.14 to 0.40) 0.35</td>
<td>0.05 (–0.22 to 0.32) 0.71</td>
<td>0.07 (–0.20 to 0.33) 0.64</td>
<td>0.11 (–0.16 to 0.38) 0.43</td>
</tr>
<tr>
<td>Normal morphology (%)</td>
<td>−1.17 (–1.42 to –0.92) &lt;0.001</td>
<td>−1.28 (–1.53 to –1.04) &lt;0.001</td>
<td>−1.29 (–1.54 to –1.05) &lt;0.001</td>
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</tr>
<tr>
<td><strong>Dichotomous (&lt;10th percentile as decreased)</strong>*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Decreased concentration</td>
<td>0.90 (0.85 to 0.96) &lt;0.001</td>
<td>0.90 (0.85 to 0.96) 0.001</td>
<td>0.90 (0.85 to 0.96) 0.001</td>
<td>0.90 (0.85 to 0.96) 0.001</td>
</tr>
<tr>
<td>Decreased total motility</td>
<td>0.90 (0.84 to 0.96) &lt;0.001</td>
<td>0.92 (0.86 to 0.98) 0.01</td>
<td>0.92 (0.86 to 0.98) 0.01</td>
<td>0.92 (0.86 to 0.99) 0.02</td>
</tr>
<tr>
<td>Decreased progressive motility</td>
<td>0.93 (0.88 to 0.99) 0.03</td>
<td>0.95 (0.89 to 1.01) 0.11</td>
<td>0.95 (0.89 to 1.01) 0.11</td>
<td>0.95 (0.90 to 1.01) 0.12</td>
</tr>
<tr>
<td>Decreased normal morphology</td>
<td>1.24 (1.18 to 1.31) &lt;0.001</td>
<td>1.26 (1.20 to 1.33) &lt;0.001</td>
<td>1.26 (1.20 to 1.33) &lt;0.001</td>
<td>1.26 (1.20 to 1.33) &lt;0.001</td>
</tr>
</tbody>
</table>

*Crude model had no adjustment; model 1 was adjusted for age, educational level, smoking status, alcohol drinking, exercise and occupational exposure to asbestos and organic solvent; model 2 was further adjusted for body mass index, systolic blood pressure, fasting blood glucose and total cholesterol levels; model 3 was further adjusted for season and year of medical examination.
†Beta-coefficients were presented for continuous variables and ORs were presented for dichotomous variables.
‡Concentration was log-transformed to achieve approximate normality for the data analysis and then the geometric coefficients were transformed back for presentation.
*Coef, coefficients; PM~2.5, particulate matter with an aerodynamic diameter <2.5 µm.

Effect estimates were calculated for an increment of 5 µg/m³ in PM~2.5 concentrations.

Environment

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PAHs, a group of compounds that includes several endocrine disruptors, can also cause sperm alterations through its effects on the hypothalamic-pituitary axis and testicular spermatogenesis. We observed a positive association between exposure and sperm concentration. This finding is not in line with previous studies, most of which have not observed significant associations between air pollution and sperm concentration. Sokol et al reported a negative association between O₃ and sperm concentration. A recent study by Zhou et al reported a positive association between sperm concentration and PM₁₀, which is similar to our study. Regarding motility, the associations also remain uncertain. Hammoud et al and Selevan et al observed a negative association, whereas no associations were observed in other studies. The reasons for the positive associations between exposure and sperm concentration in the present study need to be elucidated by further studies. We hypothesise that the slight increases in sperm concentration are a compensatory phenomenon. Numerous studies have demonstrated the hormesis phenomenon on chemical aerosol exposure and follicle-stimulating hormone (FSH) and luteinising hormone (LH). Low-dose exposure may increase the levels of FSH and LH, which strengthen spermatogenesis, thus increasing sperm concentration. Further studies are warranted to investigate the underlying mechanism.

We did not observe decline trends in semen quality over the study period. Although the trend of progressive motility was statistically significant, the figures fluctuated over time. The reported global downwards trend in semen quality has not been concluded because previous studies exhibit great heterogeneity due to geographical and/or ethnic variations and different study designs and methodological standards. In the present study, we adjusted for the year in the data analyses and the adjustment had few effects. Compared with previous studies, our study has some important strengths. The large sample size enabled us to have sufficient power to detect the small effects by low-dose exposure. Furthermore, our participants were from a general population rather than being men from infertility clinics. The affluent information on a wide range of potential confounders and modifiers enabled us to consider the effects of these factors. Another advantage is that we used accurate, high-resolution satellite-based technology to estimate PM₁₀ exposure at the individual level. All of the previous studies estimated air pollution exposure at the community level, which may mask exposure variation and result in misclassification. This technology also enables us to obtain short-term and long-term exposure for comparison.

However, there are limitations. First, all of the participants in this study were volunteers, and information on their history of infertility diseases was not available. We therefore could not exclude the possible influence of infertility disorders for some of the participants, but it should not affect our conclusions because PM₁₀ exposure was unlikely to be differentially distributed among fertile and infertile participants. In addition, we got similar results in subgroup analyses stratified by normal and abnormal semen parameters. Given that the participants were well educated, we should be cautious when generalising the findings into general populations. Second, most participants (97%) had only one semen measurement and this limited us to perform a repeated-measures analysis, which would address within-person variations over time. Another limitation is that we only estimated ambient PM₁₀ levels. Information on indoor PM₁₀ was not available. Despite evidence that indoor and outdoor PM levels are highly correlated, we cannot exclude the possible influence of indoor PM sources on some of the participants. In addition, the PM₁₀ concentrations were calculated at the fixed addresses and participants’ activity pattern was not taken into account. More advanced techniques are needed for more accurate exposure assessment. Our study also lacked data on other gaseous air pollutants and future studies need to take it into consideration. Finally, the long-term and short-term (a spermatogenic cycle) exposure are highly correlated in the present study. We are not able to distinguish the effects of short-term exposure.

CONCLUSIONS

In conclusion, we found a robust association between exposure to PM₁₀, air pollution and low percentage of sperm normal morphology in reproductive-age men. Although the effect estimates are small and the significance might be negligible in a clinical setting, this is an important public health challenge. Given the ubiquity of exposure to air pollution, a small effect size of PM₁₀ on sperm normal morphology may result in a significant number of couples with infertility. We advocate global strategies on mitigation of air pollution to improve reproductive health. In contradiction to the negative associations with morphology, we observed exposure to PM₁₀ associated with a higher level of sperm concentrations. We speculate the positive associations may be due to the compensation mechanism. Further studies are warranted to investigate this phenomenon and its underlying mechanism.

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