Excretion of cadmium through bile and intestinal wall in rats

M. CIKRT and M. TICHÝ
Institute of Hygiene and Epidemiology, Department of Industrial Hygiene and Occupational Diseases, Prague, Czechoslovakia

Cikrt, M., and Tichý, M. (1974). British Journal of Industrial Medicine, 31, 134-139. Excretion of cadmium through bile and intestinal wall in rats. The excretion of Cd²⁺ through the bile and intestinal wall after intravenous administration of CdCl₂ in non-toxic doses (67, 90, and 120 µg of Cd²⁺ per rat) was studied in rats. The cumulative biliary excretion reached 24 hours after administration of the 67 µg dose was 0.83%, after 90 µg 1.18%, and after the 120 µg dose 5.68% of the amount given. The highest excretion rate of Cd²⁺ was detected between 15 and 30 minutes after administration. There was no difference in the excretion through intestinal wall between the 67 and 120 µg doses of Cd²⁺ per rat. The mean amount of cadmium found in the contents of the entire gastrointestinal tract and faeces was 5.5% of the administered dose. Using polyacrylamide gel disc electrophoresis it was found that Cd²⁺ is bound with at least two different components of the rat bile.

In the past two decades there has been an increase in industrial production and use of cadmium. Concomitantly, there has been an increased prevalence of both acute and chronic cases of clinically identifiable cadmium poisoning (Flick, Krabyll, and Dimitroff, 1971). Cadmium has been found not only deposited and accumulated in various body tissues but also in varying concentration throughout all environmental compartments (air, water, food, and soil).

For hitherto unknown reasons cadmium is excreted very slowly by both experimental animals and man and tends to accumulate in the body (Berlin and Ullberg, 1963; Lucis, Lynk, and Lucis, 1969). After subcutaneous or intravenous administration cadmium is concentrated predominantly in the kidneys, liver (Berlin and Ullberg, 1963; Lucis et al., 1969), pancreas, spleen, and intestine (Decker and Byerrum, 1956), and even in the endocrine glands (Berlin and Ullberg, 1963). During the 10 minutes after intravenous administration the content of cadmium in liver quickly increases whereas the content of the kidneys remains relatively constant in the same period (Perry et al., 1970).

Berlin and Ullberg (1963) studied the body distribution of cadmium in mice at various times after a single intravenous dose of ¹⁰⁹CdCl₂, using the autoradiography method with sagittal whole-body sections. They found that cadmium left the blood very soon after injection and accumulated in the liver, kidneys, and mucous membranes of the intestinal tract. The concentration of cadmium in the organs did not change in course of the experiment but in the mucosa of the intestinal tract it decreased during the first 24 hours. According to these authors it seems likely that the mucosa of the intestinal tract, especially of the stomach and colon, took part in the excretion of cadmium. The autoradiograms strongly suggested excretion of cadmium in the bile.

According to Lucis et al. (1969) the principal route of cadmium excretion was via the gastrointestinal tract. These authors supposed that the appearance of cadmium in the lumen of the small intestine might be a result of biliary, pancreatic, and intestinal secretions. They believed that the colon might be involved in reabsorption of cadmium during the first 24 hours after injection.

In our study we followed the biliary excretion of
cadmium and its excretion through the wall of the gastrointestinal tract after intravenous administration of Cd\(^{2+}\) to rats. Using polyacrylamide gel disc electrophoresis we compared the protein spectrum of rat bile with the location of \(^{115}\)mCd on the electrophoreogram.

**Material and methods**

Female Wistar rats, mean weight 200 g (180-220 g) and fed on a pellet diet, were used in the experiments. Rats were starved for 24 hours before the study but had free access to water.

The bile duct was cannulated with PE-10 tubing as described by Cikrt (1972). For separation of urine and faeces we used the modified method described by Östlund (1969).

Bile samples were taken at one-hour intervals and in some cases at two-hour intervals. The total duration of the experiment was 24 hours. The rats were given intravenously CdCl\(_2\) in doses of either 67 \(\mu \text{g}\), 90 \(\mu \text{g}\) or 120 \(\mu \text{g}\) of Cd\(^{2+}\) per rat. In another series of experiments the rate of excretion of cadmium in the bile was followed for the first 80 minutes after administration. In these experiments bile was taken every 5 minutes. The rats were given \(^{115}\)mCdCl\(_2\) in doses of 95 \(\mu \text{g}\) of Cd\(^{2+}\) per rat. The radioactivity of the administered isotope amounted to 10 \(\mu \text{Ci}\) per rat.

After 24 hours (or 80 minutes respectively) the rats were decapitated; blood was collected and heparinized, and the liver, both kidneys, and the entire gastrointestinal tract were carefully removed from the abdominal cavity. Plasma was separated by centrifugation. The gastrointestinal tract was divided into six parts according to anatomical segments (Cikrt, 1972). The concentration of cadmium was determined both in the wall of the gastrointestinal tract and in the intestinal contents.

Cadmium concentration was determined with a Varian Techtron Atomic Absorption Spectrophotometer model AA-4 equipped with a Cd hollow cathode lamp. The analytical spectral line at 2290.5 Å was used. The results were not corrected for light absorption of control bile samples.

The samples of bile and plasma were diluted with deionized water to 1 ml. Tissue samples, intestinal contents, faeces, and urine collected on filter paper were ashed by a wet method using nitric acid, and after removal of excess acid the residue was dissolved in 10 ml of deionized water. In control samples of bile collected one hour before the injection of cadmium, absorption spectrophotometry with a Cd hollow cathode lamp was also carried out.

The measurement of radioactivity was carried out for \(^{115}\)mCd in a well-type scintillation counter. Measurement time was selected after radioactivity sampling had demonstrated significant differences from the background. Counts were within a ±5% error.

The number of rats in individual experiments varied from five to eight animals. The results are expressed in percentage of administered doses (means and 95% confidence limits).

For plasma the results are expressed in terms of the entire plasma volume (8.3 ml per 200 g rat (Spector, 1956)). The data in the tables for liver and kidneys are expressed as the percentage of the administered dose in relation to the entire liver or both kidneys.

The cadmium content in the wall and lumen of the individual anatomical segments of the gastrointestinal tract is given as a percentage of the administered dose. The content of cadmium in the faeces collected during 24 hours after dosing was added to that found in the colon lumen.

The bile was fractionated by the method of disc electrophoresis on polyacrylamide gel (Davis, 1964). The bile

---

**FIG. 1.** Biliary excretion of 67 \(\mu \text{g}\) Cd\(^{2+}\) per rat administered intravenously over 24 hours: (A) cumulative excretion as percentage of the administered dose; (B) percentage of excreted Cd\(^{2+}\) per milligramme of bile; (C) excretion of Cd\(^{2+}\) as percentage of the administered dose per minute; (D) bile flow in milligrammes per minute (mean ± 95% confidence limits of means). Solid lines—results from individual rats; open circles—mean values.
collected from rats, after intravenous administration of 90 μg of $^{114m}$Cd$^{2+}$, was placed on polyacrylamide gel. After electrophoresis each gel was cut into twelve 5-mm slices which were then allowed to dissolve in 0.2 ml of 30% H$_2$O$_2$ for 12 hours at 60°C; 6 ml of scintillation solution, 3 ml of absolute ethanol and 0.5 ml of hyamine hydroxide were added to the dissolved samples which were then measured with a Tri-Carb Liquid Scintillation Spectrometer (Packard, model 3365). The cadmium content of individual sections of electrophoreograms is given as the percentage of the total amount of radioactivity $^{114m}$Cd determined.

Results

The biliary excretion of cadmium in 24 hours is given as the mean and as the absolute values obtained.

![Fig. 2](image1.png)

**Fig. 2.** Biliary excretion of 90 μg Cd$^{2+}$ per rat administered intravenously over 24 hours: (A) cumulative excretion as percentage of the administered dose; (B) percentage of excreted Cd$^{2+}$ per milligramme of bile; (C) excretion of Cd$^{2+}$ as percentage of the administered dose per minute; (D) bile flow in milligrammes per minute (means ± 95% confidence limits of means). Solid lines—results from individual rats; open circles—mean values.

![Fig. 3](image2.png)

**Fig. 3.** Biliary excretion of 120 μg of Cd$^{2+}$ per rat administered intravenously over 24 hours; (A) cumulative excretion as percentage of the administered dose; (B) percentage of excreted Cd$^{2+}$ per milligramme of bile; (C) excretion of Cd$^{2+}$ as percentage of the administered dose per minute; (D) bile flow in milligrammes per minute (means ± 95% confidence limits of means). Solid line—results from individual rats; open circles—mean values.
Excretion of cadmium through bile and intestinal wall in rats from individual rats (Figs. 1 to 3). We believe that this method of presentation better expresses the uniform character of excretion than statistical characteristics only. In Figs. 4 and 5, which show the results of biliary excretion of $^{116m}$Cd within 80 minutes after dosing, only means and 95% confidence limits are shown.

**FIG. 5.** Distribution of $^{115m}$Cd$^{2+}$ detected on electrophoreogram of rat bile after intravenous administration of 90 μg of $^{115m}$Cd$^{2+}$ per rat. The values are expressed as the percentage of the total amount of radioactivity determined on the electrophoreogram. Each column in the diagram represents the mean of six individual determinations.

### Biliary excretion of cadmium

In the 24 hours experiments the following amounts of cadmium were excreted in the bile:

<table>
<thead>
<tr>
<th>Administered dose (μg Cd$^{2+}$ per rat)</th>
<th>Biliary excretion/24 hr (% of administered dose)</th>
</tr>
</thead>
<tbody>
<tr>
<td>67</td>
<td>0.83 ± 0.18</td>
</tr>
<tr>
<td>90</td>
<td>1.18 ± 0.36</td>
</tr>
<tr>
<td>120</td>
<td>5.68 ± 1.86</td>
</tr>
</tbody>
</table>

The curves B and C of Figs. 1, 2, and 3 show that the greatest part of cadmium is excreted in the bile within the first hour after its administration. In order to determine the position of the maximum rate of excretion we followed up the biliary excretion of $^{116m}$Cd for 80 minutes after dosing (Fig. 4). The highest excretion rate of $^{116m}$Cd was detected between 15 and 30 minutes after administration (Fig. 4B and C).

**FIG. 4.** Biliary excretion of 95 μg of $^{116m}$Cd$^{2+}$ per rat administered intravenously over the first 80 minutes after dosing: (A) cumulative excretion as percentage of the administered dose; (B) percentage of excreted $^{116m}$Cd$^{2+}$ per milligramme of bile; (C) excretion of $^{116m}$Cd$^{2+}$ as percentage of the administered dose per minute; (D) bile flow in milligrammes per minute. Open circles—mean values (7 rats measured); vertical lines—95% confidence limits of means.
Excretion of cadmium via wall of gastrointestinal tract

Table 1 shows the cadmium content as the percentage of the administered doses for the entire alimentary tract found in the lumen of the individual segments 24 hours after administration. In the table the results of the experiment in which 90 μg of Cd\(^{2+}\) were administered are not presented.

It will be seen that the largest amount of metal was found in the lumen of the caecum and colon including faeces. The total amount of the metal excreted within 24 hours via the wall of the gastrointestinal tract did not differ in the two doses of Cd\(^{2+}\) given.

### TABLE 1

**Cadmium Content in Lumen of Gastrointestinal Tract 24 Hours after Administration**

<table>
<thead>
<tr>
<th>Dose (μg per rat)</th>
<th>67</th>
<th>120</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stomach</td>
<td>0.34 ± 0.15</td>
<td>0.22 ± 0.11</td>
</tr>
<tr>
<td>Duodenum</td>
<td>0.19 ± 0.08</td>
<td>0.30 ± 0.07</td>
</tr>
<tr>
<td>Jejunum</td>
<td>0.26 ± 0.10</td>
<td>0.56 ± 0.37</td>
</tr>
<tr>
<td>Ileum</td>
<td>0.36 ± 0.04</td>
<td>0.41 ± 0.25</td>
</tr>
<tr>
<td>Caecum</td>
<td>2.63 ± 0.50</td>
<td>2.51 ± 0.81</td>
</tr>
<tr>
<td>Colon + faeces</td>
<td>1.52 ± 0.34</td>
<td>1.68 ± 0.25</td>
</tr>
<tr>
<td>Entire gastrointestinal tract + faeces</td>
<td>5.31 ± 1.21</td>
<td>5.68 ± 1.86</td>
</tr>
</tbody>
</table>

The values given in the table are means and 95% confidence limits of means. Values are expressed as a percentage of the administered dose.

Content of cadmium in liver, kidneys, urine, plasma, and wall of gastrointestinal tract

Table 2 shows the Cd\(^{2+}\) content in liver, kidneys, urine, and plasma 24 hours after dosing. It is apparent that there are no significant differences in the cadmium content of various tissues after different Cd\(^{2+}\) doses.

### TABLE 2

**Cadmium Content in Liver, Kidneys, Plasma, and Urine 24 Hours after Administration**

<table>
<thead>
<tr>
<th>Dose (μg per rat)</th>
<th>Liver</th>
<th>Kidneys</th>
<th>Plasma (8.3 ml)</th>
<th>Urine (per 24 hr)</th>
</tr>
</thead>
<tbody>
<tr>
<td>67</td>
<td>79 ± 1 ± 3.1</td>
<td>2.3 ± 0.2</td>
<td>0.51 ± 0.06</td>
<td>1.7 ± 0.9</td>
</tr>
<tr>
<td>90</td>
<td>76 ± 3 ± 3.4</td>
<td>2.7 ± 0.2</td>
<td>*</td>
<td>0.9 ± 0.1</td>
</tr>
<tr>
<td>120</td>
<td>67 ± 3 ± 8.5</td>
<td>1.9 ± 0.5</td>
<td>0.25 ± 0.02</td>
<td>0.52 ± 0.14</td>
</tr>
</tbody>
</table>

*Not analysed

Values are expressed as a percentage of the administered dose (means and 95% confidence limits of means).

Table 3 shows the cadmium content in the wall of the individual anatomical segments of the gastrointestinal tract 24 hours after administration.

### TABLE 3

**Cadmium Content in Wall of Gastrointestinal Tract 24 Hours after Administration**

<table>
<thead>
<tr>
<th>Dose (μg per rat)</th>
<th>67</th>
<th>120</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stomach</td>
<td>0.74 ± 0.16</td>
<td>0.60 ± 0.12</td>
</tr>
<tr>
<td>Jejunum</td>
<td>0.18 ± 0.43</td>
<td>0.85 ± 0.14</td>
</tr>
<tr>
<td>Ileum</td>
<td>0.52 ± 0.70</td>
<td>0.01 ± 0.22</td>
</tr>
<tr>
<td>Caecum</td>
<td>0.44 ± 0.35</td>
<td>0.96 ± 0.20</td>
</tr>
<tr>
<td>Colon</td>
<td>0.62 ± 0.10</td>
<td>0.49 ± 0.05</td>
</tr>
<tr>
<td>Entire gastrointestinal tract</td>
<td>5.4 ± 0.68</td>
<td>0.88 ± 0.18</td>
</tr>
<tr>
<td>7.04 ± 2.42</td>
<td>4.79 ± 0.91</td>
<td></td>
</tr>
</tbody>
</table>

Values are expressed as a percentage of the administered dose (means and 95% confidence limits of means).

In the experiments involving administration of 95 μg of \(^{115m}\text{Cd}^{2+}\) per rat we found, 80 minutes after dosing, in the liver 72.2 ± 0.3%, in both kidneys 1.84 ± 0.03%, and in the plasma 3.59 ± 0.25% of the administered dose.

The results for bile, kidneys, and liver obtained by atomic absorption spectrophotometry were compared with the data obtained using radioisotope \(^{115}\text{Cd}\). No significant differences were observed. Moreover, the cadmium content in the intestinal lumen and faeces after the first 24 hours was no higher in our experiment than the values found by Decker et al. (1957). These authors used \(^{115}\text{Cd}\) and similar experimental conditions. The differences are within the usual range of biological variation.

Fractionation of bile by disc electrophoresis on polyacrylamide gel

The protein electrophoretic spectrum of the bile of rats given \(^{115m}\text{CdCl}_2\) intravenously did not differ from the protein electrophoretic spectrum of the control bile taken from the same animals before administration of the radioisotope. The distribution of radioactivity of the protein electrophoretic spectrum of this bile is shown in Figure 5. Cd\(^{2+}\) cations are concentrated in two parts of the electrophoreogram —in the pigment zone, and between the prestacking gel and the midzone (Englert, Wales, and Stradley, 1970). Between both maxima, a part containing a smaller amount of cadmium was found.

**Discussion**

The biliary excretion of cadmium in the 24 hours after administration is very low. After the dose of 120
μg Cd²⁺ per rat we observed a higher biliary excretion of cadmium than after administration of 67 μg or 90 μg Cd²⁺ per rat. The highest biliary excretion of cadmium per minute is reached between 15 and 30 minutes after dosing.

Berlin and Ullberg (1963) and Lucis et al. (1969) observed that there was a possibility that cadmium was excreted in the bile. In our experiments less cadmium is excreted in the bile than via the wall of the gastrointestinal tract. Berlin and Ullberg (1963) suppose that cadmium is excreted into the faeces, particularly by the gastric mucosa and the mucosa of the colon, within the first 24 hours after administration. Lucis et al. (1969), however, emphasize the importance of the excretion of cadmium into the small intestine with subsequent possible reabsorption in the colon. In our experiments we found a relatively higher content of cadmium in the wall of the entire gastrointestinal tract by comparison with several other metals (Cikrt, 1972). The largest amount of cadmium in the intestinal content was found in the caecum and colon, including the faeces. The transit time of the intestinal contents is relatively rapid in the duodenum and upper jejunum and is reduced in the distal parts of the gut (Marcus and Lengemann, 1962). In the caecum the food remains for a long period; thus it may be presumed that the high content of cadmium found in the lumen of the caecum is due to the accumulation of the metal excreted in the upper segments of the digestive tract. Our results support the view that the colon might also play an important part in cadmium excretion. This observation would be in conformity with the autoradiographical findings of Berlin and Ullberg (1963). Our results are in good agreement with the data of Decker, Byerrum, and Hoppert (1957), who found 73% of cadmium in the faeces 24 hours after the administration of a single intravenous dose of 115Cd (0.63 mg per kg) to rats.

We have shown by electrophoretic fractionation of bile that cadmium is bound with at least two different components of bile. Part of the cadmium is accumulated in the pigment zone and another part in the region between the prestacking gel and midzone. Both parts are of practically equal size. Examination of bile by a chromatographic fractionation method using a Sephadex G-100 column (Havrdoval, Cikrt, and Tichy, to be published) gave similar results. Part of the cadmium was bound to low molecular weight components and part to high molecular weight components of the bile. It is not certain whether the low molecular weight component is identical with metallothionein. It is known, however, that the formation of metallothionein in the liver is induced after administration of cadmium (Piscator, 1964). Because of the significant porosity of the hepatobiliary system (Schanker and Hogben, 1961) it is possible that transport of cadmium bound to metallothionein into bile takes place.

References


Received for publication 20 February 1973
Accepted for publication 6 November 1973