Effect of phenobarbitone and starvation on hepatotoxicity in rats exposed to carbon disulphide vapour

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Carbon disulphide (CS₂) is not recognized as an hepatotoxic agent. Rabbits exposed daily—five days a week—for many weeks until a partial loss of control over voluntary movements of the hind legs appeared did not show any pathological changes in the liver (Cohen et al., 1959). An oral dose of 1 mg/kg CS₂ given to rats produced only an increase in fat in the perportal zone. When the same dose of CS₂ was given to rats pretreated with phenobarbitone, extensive centrlobular zone necrosis appeared (Bond, Butler, De Matteis, and Barnes, 1969). However, these animals were subjected not only to phenobarbitone pretreatment but also to starvation as food was withdrawn 24 hours before exposure to avoid the gastric effect of CS₂ on food and food consumption.

The purpose of the present work was to investigate whether liver damage can be induced in phenobarbitone-pretreated rats by inhalation exposure to CS₂. Inhalation exposure also made it possible to study the effect of starvation on liver damage in phenobarbitone-pretreated rats exposed to CS₂. Starvation significantly decreased the urinary excretion of both CS₂ and its bivalent sulphur metabolites in exposed rats (Magos, 1972) and starvation may have contributed to the development of liver necrosis in phenobarbitone-pretreated animals (Bond et al., 1969). As CS₂ can be converted in the body to dithiocarbamate type compounds (Souček, 1957) and dithiocarbamate is partly metabolized to CS₃ (Strömme, 1965) the effect of diethylthiocarbamate on the liver was also investigated in some of the experiments.

Methods

Male albino rats of Porton-Wistar strain (200-220 g) bred in this colony were used. Phenobarbitone sodium (BDM) was given intraperitoneally in a dose of 80 mg/kg 24 hours and 50 mg/kg 18 hours before 4-hour exposure to 2 mg/kg CS₂ or 18 and 23 hours before the administration of 500 mg/kg sodium diethylthiocarbamate (DDC; Hopkin and Williams). Animals were exposed to CS₂ in a vertical type inhalation chamber (Magos, Emery, Lock, and Firmager, 1970). From the starved group, food was withdrawn 24 hours before exposure or before DDC. From the fed animals, food was withdrawn at the beginning of exposure to CS₂ or 5 hours before the injection of DDC. Sucrose or fat fed animals were given only sucrose or only pork lard in the last 24 hours before exposure. Water was available ad libitum to all animals except when they were in the inhalation chamber.
chamber. Animals were killed by decapitation, usually 18 hours after the end of exposure or the administration of DDC, and the livers were removed, weighed, and fixed in formol-alcohol or formol saline. Paraffin sections prepared in the usual fashion were stained with Harris' haematoxylin and eosin, and in some cases frozen sections were stained with oil red 0 for fat.

The liver sections were examined without knowledge of the treatment and were given a score indicating the severity of the damage. Sections scored as 0 were indistinguishable from normal. A score of 1 indicated a few hydropic cells in the region of the central vein (Fig. 1) while a score of 2 indicated that in the order of half of each lobule was changed to hydropic cells (Fig. 2). A score of 3 was used when only a rim of hepatic parenchymal cells remained in the vicinity of the portal tracts surrounding the more centrilobular hydropic cells.

Results

The character of the histological change at 18 hours was similar in all the rats treated with phenobarbitone and exposed to CS₂ but the extent of the damage was variable. The main change in the hepatic parenchymal cells of the centrilobular area was a swelling of the cytoplasm similar to that of a hydropic degeneration with the nuclei of those cells normal in appearance. However, scattered throughout the zone of hydropic degeneration were pyknotic nuclei, and nuclei in which there was margination of the chromatin or fragmentation of the nucleus. The central veins were normal and no inflammatory reaction was seen. In the periploral zone the surviving hepatic cells were normal except for an accumulation of some lipid, not seen in the centrilobular area, and slightly greater than that seen in the unexposed starved animals with or without phenobarbitone treatment. The portal tracts were normal. Within any liver the extent of the change was reasonably uniform.

Table 1 shows the degree of liver damage, scored as described above, produced by 4-hour exposure to CS₂ (2.0 mg/l) in phenobarbitone-pretreated rats and that phenobarbitone was essential for the development of liver damage in starved rats exposed to CS₂ or treated with diethyldithiocarbamate. When 500 mg/kg DDC was given to phenobarbitone-pretreated rats before exposure to CS₂ (group 5) the liver damage was more severe than in either CS₂-exposed (group 2) or DDC-treated rats (group 4). The extent of liver damage in the DDC-treated rats (group 4) was less severe than in CS₂-exposed rats (group 2). There is some variation within groups as indicated by the scoring of the histological

![Fig. 1](http://oem.bmj.com/)

**Fig. 1.** Liver of rat, pretreated with phenobarbitone and starved, killed 24 hours after exposure to carbon disulphide showing a centrilobular zone of hydropic degeneration. Grade 1 lesion. H. and E. × 130.
Effect of phenobarbitone and starvation on hepatotoxicity in carbon disulphide exposed rats

FIG. 2. Liver of rat, pretreated with phenobarbitone and starved, killed 24 hours after exposure to carbon disulphide showing extensive hydropic degeneration involving about one-half of the lobule. Grade 2 lesion. H. and E. × 130.

TABLE 1

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>No. of rats</th>
<th>Grade of liver damage</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Ph DDC CS₂</td>
<td>0 1 2 3</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>- - +</td>
<td>16</td>
<td>16 - - -</td>
</tr>
<tr>
<td>2</td>
<td>+ - +</td>
<td>6</td>
<td>- 1 5 -</td>
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<tr>
<td>3</td>
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</tr>
<tr>
<td>4</td>
<td>+ + +</td>
<td>6</td>
<td>1 4 1 -</td>
</tr>
<tr>
<td>5</td>
<td>+ + +</td>
<td>6</td>
<td>- - 1 5</td>
</tr>
<tr>
<td>6</td>
<td>- - -</td>
<td>8</td>
<td>8 - - -</td>
</tr>
</tbody>
</table>

Phenobarbitone sodium (Ph) in a dose of 80 and 50 mg/kg was given 24 and 18 hours before exposure or the administration of DDC. Food was withdrawn 24 hours before exposure. Animals were killed 24 hours after injection or the beginning of exposure.

lesions. The differences between the severity of the lesions as illustrated in Figs. 1 and 2 is such that in spite of the difficulties in scoring degrees of damage these groups are distinct.

Feeding provided a partial protection against the hepatotoxic effect of CS₂ in phenobarbitone-pretreated rats. Only 10 of the 30 fed animals developed any discernible liver lesions compared with 28 of the 30 starved rats (Table 2). When incomplete diets were given in the form of sucrose or animal fat, the frequency and severity of the lesions were somewhat between those observed in starved and fed animals.

One-hour CS₂ exposure produced liver lesions in five of the eight starved, phenobarbitone-treated rats killed 21 hours after exposure, though in four animals damage in the liver scored only 1+. There was no difference in liver damage between animals exposed 3 or 4 hours to 2-0 mg/l CS₂. An interval between exposure and sacrifice was essential for the development of the hepatic lesion as none of the five starved, phenobarbitone-pretreated rats killed immediately after exposure showed signs of liver damage.
TABLE 2

**Effect of Starvation or Incomplete Feeding on Liver Damage in Phenobarbitone-Pretreated Male Rats exposed 4 hours to 2-0 mg/l CS2**

<table>
<thead>
<tr>
<th>Nutritional condition</th>
<th>No. of rats</th>
<th>Grade of liver damage</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>0</td>
</tr>
<tr>
<td>Fed</td>
<td>30</td>
<td>20</td>
</tr>
<tr>
<td>Fasted</td>
<td>30</td>
<td>2</td>
</tr>
<tr>
<td>Sucrose-fed</td>
<td>15</td>
<td>2</td>
</tr>
<tr>
<td>Fat-fed</td>
<td>6</td>
<td>1</td>
</tr>
</tbody>
</table>

Food was withdrawn 24 hours before exposure when the first phenobarbitone dose (80 mg/kg) was given; 6 hours later phenobarbitone treatment was repeated by giving a 50 mg/kg dose. Rats were killed 24 hours after the beginning of exposure.

Discussion

We have confirmed the observation that CS2 can only induce liver damage in male rats which have been pretreated with phenobarbitone but the dose by inhalation is considerably less than the 1.0 ml/kg per os given (Bond et al., 1969). In rats exposed to 2-0 mg/l CS2 for 4 hours, even the all the CS2 which reached the alveoli to be taken up by the circulating blood, the amount absorbed can be no more than 100 mg/kg. Nevertheless the histological changes in the starved animals exposed to vapour are comparable to those in rats given CS2 per os.

The toxic reaction in the liver can only be demonstrated when exposure follows phenobarbitone treatment; the reaction is aggravated by starvation. The observation that starvation increases the effect of phenobarbitone on cytochrome P-450 concentration in the liver (Greim, 1970) offers a very attractive hypothesis for the explanation of phenobarbitone effect on CS2 hepatotoxicity. Phenobarbitone might increase the transformation of CS2 to a toxic metabolite through the induction of drug-metabolizing enzymes, as has been suggested for CCl4, a rather unreactive compound. However, this hypothesis presents some difficulties. Compared with carbon tetrachloride, CS2 is an extremely reactive compound and there is no evidence of the existence of a more reactive metabolite. Furthermore, the urinary excretion of CS2 and its bivalent metabolites is not altered by phenobarbitone treatment but only by starvation (Magos, 1972). Finally, in agreement with the metabolic pathway of DDC, which is mainly detoxified by conjugation with glucuronic acid and to a smaller extent decomposed to CS4 (Strömme, 1965), DDC is able to produce an hepatotoxic effect in fasted phenobarbitone-treated rats, but even a 500 mg/kg dose resulted in only slight changes compared with those observed in rats exposed to CS2. Consequently, it seems worthwhile to consider the possibility that CS4 and not one of its metabolites is the hepatotoxic agent and that starvation aggravates hepatotoxicity by promoting the retention of CS4 or its additive reaction with cell constituents.

The practical implications of these experiments are that in certain conditions of exposure to the vapours of CS2 hepatic damage may result. Thus the effects of an occupational exposure dose might be changed by factors such as the nutritional state or the administration of therapeutic drugs. Though the CS2 concentration was very high in these experiments compared with the maximum allowable concentration (0-06 mg/l), one must consider that on the one hand the exposure time was extremely short and on the other hand much higher concentrations than the MAC value have been reported from industry (Brieger and Teisinger, 1967).

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References


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