Some peculiarities of the pulmonary phagocytotic response: dust retention kinetics and silicosis development during long term exposure of rats to high quartz dust levels

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ABSTRACT Rats were exposed to quartz dust (about 90 mg/m³) for five hours a day, five times a week either throughout the 48 weeks of the experiment or for a total of 40 weeks plus eight weeks of “rest.” Cytological study of bronchoalveolar lavage showed that at a certain level of silicotic changes in the lungs, a pronounced breakdown in pulmonary dust clearance by macrophages could be observed. There was, however, a concomitant compensatory increase in the contribution to pulmonary phagocytosis by the neutrophil leukocytes (NL). As a result, pulmonary dust kinetics could be mathematically described using a constant clearance rate both throughout the 48 weeks of exposure and during the eight weeks after the end of the 40 week exposure. It is shown in a separate experiment that enhanced recruitment of NL into the airways may be due not only to an attractant effect of the products of macrophage breakdown (PMB) themselves but also to the release of an NL attractant factor by viable macrophages activated under influence of the PMB.

It is beyond doubt that recruitment of phagocytosing cells to the free surface of the lower airways plays a key part in the elimination of deposited particles to the zone of efficient mucociliary transport and thus precludes their penetration into fixed lung tissues and translocation to lymph nodes. It was shown earlier that not only alveolar macrophages (AM) but neutrophil leukocytes (NL) contribute significantly to dust elimination and that recruitment of both cell types is regulated, first of all, through a complex self regulating mechanism. The main feature of the latter is its dependence on the amount of lipid containing products of macrophage breakdown (PMB), this in turn depending on the level of dust exposure and the cytotoxicity of the dust particles. This self regulation, however, acts in accordance with the host’s general reactivity and is influenced by neurohormonal regulation.

Whether alterations in the operation of the phagocytotic mechanism of pulmonary dust clearance occur after long term exposure to dust is far less well known, nor is it known to what degree the kinetics of pulmonary dust retention may be changed as the result of chronic damage to this mechanism. Having found no differences in kinetics of quartz dust retention in lungs after long term exposure to different dust levels (1-0 v 20-0 mg/m³), Stöber et al nevertheless admitted the possibility of a breakdown in the pulmonary clearance mechanisms with a sufficiently large dust burden. A team of investigators from the Edinburgh Institute of Occupational Medicine, having studied the kinetics of pulmonary retention of amosite fibres, supposed that exactly such a breakdown was observed at a certain level of retention and connected it with a “dust overload” of the lungs. They expressed the opinion that it was the macrophageal mechanism of pulmonary dust clearance that suffered from such an overload.

We became interested in the question of whether the pulmonary phagocytotic response to a continuous dust exposure at a constant level really undergoes any unfavourable changes and whether there are any shifts in the dust retention kinetics corresponding to such changes. It seemed natural to study this problem, first of all, in relation to silica dust, which exerts the most prominent damaging influence on the macrophage, and to provide both a high level of dust exposure and a sufficiently long (in terms of the lifespan of the rat) exposure period.

Accepted 13 May 1986
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As the results of such experiments showed once more the most important role of the recruitment to the airways of NLs, we consider here also some new experimental data concerning the mechanisms controlling this recruitment.

Materials and methods

The inhalation experiment was carried out on male white rats with an original body weight of about 160 g. They were exposed to dust generated by milled quartzite rock from one of the Urals deposits which was more than 95% alpha-quartz. Samples of airborne dust from the exposure chambers had, on average, a particle size distribution (under light microscopy with magnification ×900) as follows: 0-1 μm: 51-3%, >1-2 μm: 33-1%, >2-5 μm: 12-0%, >5 μm: 3-6%. The animals were exposed for five hours a day, five times a week for 48 weeks, the average dust concentration for this period being 87-8 ± 3-4 mg/m³ (mean ± SE). Some 10-11 rats from the exposed and control groups were killed after 10, 20, and 40 weeks from the start of the experiment; the average dust concentrations for these periods were 87-3 ± 3-8, 84-9 ± 6-3, and 85-4 ± 3-7 mg/m³ respectively. Moreover, in the same dust chambers were rats for whom the total exposure was only 40 weeks, with three different patterns of distribution in the period between the beginning and the end of the 48 week experiment: (a) first 40 weeks of exposure and then eight weeks out of the dust chamber (the rest); (b) 20 weeks of exposure, then eight weeks of rest and 20 weeks more of exposure; (c) four subperiods of 12 weeks each comprising 10 weeks of exposure and two weeks of rest, which two weeks completed each of the first three subperiods, whereas in the fourth the rest was given in the 10th and 11th weeks and the exposure was resumed during the 12th week. The mean dust concentrations for the total 40 weeks of exposure were, respectively, 85-4 ± 3-7, 89-7 ± 4-0, and 87-5 ± 3-8 mg/m³.

Forty eight weeks after the beginning of the experiment all surviving rats were simultaneously killed (in each of four exposed groups as well as in the unexposed ones there were 18–21 surviving animals). Both in these rats and in those killed at the above mentioned intermediate stages of the weekly exposure period we determined

1. the total cell count of single bronchoalveolar lavage (BAL) according to LaBelle and Brieger with differential counting of cells in smears of centrifuged BAL sediments;
2. the dry weight of the lungs and the wet and dry weights of the tracheobronchial (TB) lymph nodes;
3. total lipid, total hydroxyproline, and dust content of the lungs.

Partly to eliminate the variability of the indices connected with the variability of body weight, not only the weight of the organs but also their lipid and hydroxyproline content were related to 100 g body weight.8

The lungs and TB lymph nodes from several rats killed at the end of the experiment were taken for histological study under the light microscope using a haematoxylin-eosin and van Gieson stains.

An additional experiment was performed on female Wistar rats to test the part taken by the activation of viable macrophages with PMB in the attraction of NLs to the lower airways. Macrophages were obtained from the peritoneal cavity of rats 48 hours after intraperitoneal injection of sterile mineral oil. PMB were produced with a triple freezing-thawing procedure as described earlier.1 Peritoneal macrophages having 97% viability in the trypan-blue exclusion test were incubated for three hours at 37°C (a) in 199 medium or (b) in the same medium plus either PMB in a dose corresponding to 2·10⁶ broken down cells per ml, or zymosan in final concentration 0·05 mg/ml. In a separate sample of 199 medium, PMB without viable macrophages were similarly incubated. Each sample was then centrifuged at 1500 rpm for 30 minutes at 4°C, and 1 ml of the supernatant was instilled intratracheally into a rat. From eight to 10 recipient rats were given supernatants obtained after each variant of the incubation or control medium.

A single bronchoalveolar lavage was performed under hexobarbitone anaesthesia after 24 hours for total and differential cell counts as mentioned above.

Results

Histological investigation of the lungs showed, in rats exposed throughout the 48 weeks, typical silicotic nodules up to the second maturity stage and also large foci of alveolar lipoproteinosis with pronounced collagen fibrosis of adjacent interalveolar septa; pronounced hyperplasia of peribronchial lymph follicles with the development of focal reticuloendothelial cell hyperplasia (this was also found in TB lymph nodes); goblet cell hyperplasia in bronchial mucosa. Lungs of rats exposed uninterruptedly during the first 40 weeks and killed after eight weeks of rest showed virtually the same histological picture. When the exposure period had been divided into four equal parts each comprising 10 weeks of exposure and two weeks of rest, silicotic nodules of the first stage predominated. Still less pronounced were the nodular lesions (with complete absence of silicotic nodules in some rats’ lungs) in the group which had been given eight weeks rest in the middle of the exposure period.

The group averages of all indices mentioned in the
materials and methods section are presented in tables 1-5 and in the figure.

Discussion

As clearly seen from table 1, recruitment of AMs to the airways changed considerably during the 48 weeks of dust exposure. If the difference between AM counts in BAL of exposed and control rats is taken as a measure of the response to the deposition of inhaled silica particles this response appears to be vigorous only in the first half of the period. At the end of the 10th week the difference was 1.45 ± 0.32 × 10^6 (p < 0.05), and at the end of the 20th week it was still greater: 3.15 ± 0.62 × 10^6 (p < 0.05). After 20 more weeks of exposure, however, this difference disappeared completely, while at the end of the 48th week it was small and statistically non-significant (0.30 ± 0.42 × 10^6). The decrease between the 20th and 48th weeks of exposure is highly significant (p < 0.01). These results seem to corroborate the hypothesis that there is a breakdown in the macrophageal pulmonary dust clearance mechanism.

As seen in the figure, the pulmonary dust retention kinetics may be approximated to a monoexponential function (\( y = 21 - 17e^{-0.0783t} \)), where “t” is time in weeks). In other words, the region of the respiratory tract where this long term retention takes place might be described using a one compartment model with constant rates of deposition (1.6443 mg a week) and of elimination (k = 0.0783 wk^-1). Such a conclusion accords well with results of other experiments with quartz dust inhalation, and closely approximates calculated model parameters. In experiments conducted on rats exposed to dust levels of either 1.0 or 20.0 mg/m^3 Stöber et al found that the pulmonary region clearance rate was, respectively, 7.4 × 10^{-3} and 9.0 × 10^{-3} for one exposure—that is, 0.0518 and 0.063 wk^-1. Taking into account the inevitable variability of biological factors (conditions of housing and feeding, age difference, and so on) and probable differences between quartz samples used in different experiments, the latter two estimates differ little from ours. Moreover, the trend of the values suggests an increase in the clearance rate with an increase in dust exposure levels.

In our experiment the efficiency of pulmonary dust clearance also remained constant in the postexposure period despite the obvious complete failure of the macrophageal clearance mechanism at the end of 40 weeks exposure. Indeed, a group of rats was allowed to "rest" after such exposure and was killed eight weeks later. Dust retention to the end of the 40th week computed from the above mentioned equation (20.25 mg) is close to the value actually found in the lungs of rats killed at that time (19.4 ± 1.7 mg) and may therefore be used as the starting lung burden (x_0) for calculating dust elimination during the eight postexposure weeks according to the equation \( Y = x_0 e^{-kt} \). If we assume that the constant clearance rate is the same as in the period of dust accumulation (k = 0.0783 wk^-1), the residual amount of dust in the lungs would be 10.82 mg. This value lies near the lower 95% confidence limit of the actual mean dust content of lungs in rats from the group surviving eight weeks after 40 weeks exposure (12.7 ± 1.0 mg, 95% confidence limits being 10.6-14.8 mg). This should be considered as a satisfactory confirmation of the mathematical prediction and, at any rate, does not confirm the hypothesis according to which, postexposure elimination of dust accumulated in lungs

<table>
<thead>
<tr>
<th>Duration of experiment (weeks)</th>
<th>Group of rats</th>
<th>No of cells in BAL × 10^6</th>
<th>Coefficient NL/AM</th>
<th>Exposed to control ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Exposed</td>
<td>Total</td>
<td>Alveolar macrophages (AM)</td>
</tr>
<tr>
<td>10</td>
<td>Control</td>
<td>1.64 ± 0.16</td>
<td>3.3 ± 0.34</td>
<td>1.26 ± 0.13</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>2.71 ± 0.29*</td>
<td></td>
<td>0.39 ± 0.09*</td>
</tr>
<tr>
<td>20</td>
<td>Control</td>
<td>1.91 ± 0.20</td>
<td>6.95 ± 0.86$</td>
<td>1.62 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>4.77 ± 0.62$</td>
<td></td>
<td>1.50 ± 0.30$</td>
</tr>
<tr>
<td>40</td>
<td>Control</td>
<td>3.22 ± 0.43</td>
<td>7.65 ± 1.05%</td>
<td>2.59 ± 0.36</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>2.58 ± 0.39</td>
<td></td>
<td>4.66 ± 0.67%</td>
</tr>
<tr>
<td>48</td>
<td>Control</td>
<td>2.48 ± 0.22</td>
<td>8.02 ± 1.00%</td>
<td>1.99 ± 0.19</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>2.29 ± 0.37</td>
<td></td>
<td>5.06 ± 0.68%</td>
</tr>
</tbody>
</table>

* and \* Values of exposed group which differ with statistical significance (with p, correspondingly, <0.05, <0.01, and <0.001) from those of controls.

\$Statistically significant difference between a given value for the exposed group and a corresponding value for the exposed rats killed at the preceding moment (p < 0.05).

\|Values significantly (p < 0.05) different from corresponding values of the 10 week exposure group.

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is considerably slowed down as a result of a long term retention.

Thus for describing the kinetics both of quartz dust accumulation in the lungs of the rat and of its subsequent clearance, even when exposure and post-exposure periods are long enough and the exposure level is high, one obviously does not need to resort either to an "overload" model or to a "sequestration" model as proposed for describing the pulmonary kinetics of asbestos. Nevertheless, how can one reconcile such a conclusion with clear evidence of a deficiency in the recruitment of AMs?

As is evident from the table 1, the recruitment of NLs to the airways of rats exposed to quartz dust during 48 weeks steadily increased. This was shown by the increase in the absolute NL count and in the NL/AM ratio. In control rats this ratio, although not increasing at the same rate, was higher in the second half of the experimental period than during the first 20 weeks. Therefore we used an additional index: the ratio of the NL/AM coefficient in an exposed group to that in the corresponding control. This index also increased throughout the 48 week exposure period.

We have already shown that the enhanced recruitment of NLs by a cytotoxic dust, or by an imitation of its action—for example, by combining exposure to particles of low cytotoxicity with intratracheal instillation of PMB—ensures the phagocytosis of a large number of particles comparable with, and sometimes even equal to, that of particles engulfed by AMs. We repeatedly emphasised in this connection that the NL response of the lower airways to the deposition of silica and other cytotoxic particles must be regarded not simply as a pathological reaction ("inflammation") but as an important compensatory one, maintaining the high efficiency of pulmonary clearance. We emphasised that this auxiliary clearance mechanism is resorted to when the main clearance mechanism (AMs recruitment and phagocytosis) is damaged, and operates as a dose dependent response to the products of AM breakdown. The results of the present experiment seem to corroborate this concept and, moreover, show that the compensation under discussion may be adequate. In fact, it would be difficult to propose another explanation for the persistence of a constant dust clearance rate against the background of the considerable damage caused to the AMs role in pulmonary phagocytosis.

There are several arguments against considering both this damage and the compensatory increase in the NLs' contribution to the phagocytotic response of the airways as a direct consequence of pulmonary dust overload. Indeed the amount of dust retained in lung tissue increased most during the first half of the exposure period (cf table 2 and the figure), whereas recruitment of AMs not only did not stop between the

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Table 2. Some quantitative indices of the state of lungs and TB lymph nodes of rats exposed for up to 48 weeks to quartz dust inhalation (five hours a day, five times a week) and of control rats (X ± Sx)

| Duration of experiment (weeks) | Group of rats | Dry weight of lungs (mg/100 g body weight) | Dry weight of TB lymph nodes (mg/100 g body weight) | Lungs
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Hydroxyproline content (µg/100 g body weight)</td>
<td>Lipid content (µg/100 g body weight)</td>
<td>Dust content (mg)</td>
</tr>
<tr>
<td>0</td>
<td>Control</td>
<td>110-8 ± 10-0</td>
<td>6-5 ± 1-6</td>
<td>1316 ± 128</td>
<td>11-3 ± 0-6</td>
<td>3-8 ± 0-5</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>107-4 ± 4-9</td>
<td>9-2 ± 1-6</td>
<td>1317 ± 73</td>
<td>11-7 ± 0-6</td>
<td>13-7 ± 1-7*</td>
</tr>
<tr>
<td>20</td>
<td>Control</td>
<td>95-4 ± 5-9</td>
<td>7-1 ± 0-6</td>
<td>1128 ± 57</td>
<td>11-1 ± 0-5</td>
<td>4-1 ± 0-6</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>177-0 ± 10-2§</td>
<td>27-1 ± 2-4§</td>
<td>1592 ± 89§</td>
<td>44-9 ± 4-0§</td>
<td>16-9 ± 2-2*</td>
</tr>
<tr>
<td>48</td>
<td>Control</td>
<td>110-0 ± 5-9</td>
<td>6-1 ± 0-7</td>
<td>949 ± 80</td>
<td>14-1 ± 0-6</td>
<td>7-4 ± 1-2§</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>224-9 ± 15-7§</td>
<td>30-8 ± 1-5§</td>
<td>2114 ± 204§</td>
<td>52-0 ± 3-2§</td>
<td>19-4 ± 1-7*</td>
</tr>
<tr>
<td>48</td>
<td>Control</td>
<td>98-6 ± 4-1</td>
<td>7-1 ± 0-5</td>
<td>1010 ± 36</td>
<td>12-9 ± 0-6</td>
<td>5-7 ± 0-8</td>
</tr>
<tr>
<td></td>
<td>Exposed</td>
<td>275-8 ± 10-5§</td>
<td>39-2 ± 1-2§</td>
<td>2426 ± 106§</td>
<td>67-2 ± 0-4§</td>
<td>20-8 ± 3-4*</td>
</tr>
</tbody>
</table>

Notation of statistical significance as in table 1.
10th and 20th weeks but was increasing at a greater rate. On the other hand, the most complete break-
down of the macrophage clearance mechanism occurred between the 20th and 40th weeks, when pul-
monary dust retention had reached its equilibrium level according to both theoretical considerations (cf
the curve shown in the figure) and experimental data (cf the figure and table 2). Moreover, the recruitment
of NLs increased up to the end of the exposure period, although the dust overload of the lungs remained
the same as eight weeks earlier.

We suppose that all the shifts in the BAL cell counts we observed are due to the development of a pul-
monary pathological process—silicosis. The severity of the latter, in its turn, depends not only on the mass
of accumulated dust (pulmonary dust burden) but also on the time of its action on the lung tissues. We
have shown that as a fairly adequate criterion for the mathematical prediction of this severity one may use
the value of the definite integral of a function describing pulmonary dust retention, the limits of integration
being the onset of exposure and any chosen moment for which the prediction is made.11 Below we shall refer
to this criterion as the “action integral” (AI).

As seen in table 2, the weight of the lungs and their lipid and hydroxyproline contents in the exposed
group did not differ from those in the control group up to the 10th week. Thus it would be justifiable to
consider the shift of BAL cellularity observed to the end of the 10th week as a “normal” defensive response
of virtually healthy lungs to the deposition of highly cytotoxic dust particles. Ten weeks later, however, the
lungs were clearly silicotic and after 40 weeks of exposure the pathological changes were even more
pronounced. It was then that the cellular composition of the BAL was most dramatically changed. At the
end of the 48 week exposure period, the severity of the

<table>
<thead>
<tr>
<th>Exposure pattern</th>
<th>No of cells in BAL × 10⁶</th>
<th>Coefficient NL/AM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total</td>
<td>Alveolar macrophages (AM)</td>
</tr>
<tr>
<td>No exposure (control)</td>
<td>2.48 ± 0.22</td>
<td>1.99 ± 0.19</td>
</tr>
<tr>
<td>48 weeks of constant exposure</td>
<td>8.02 ± 1.00*</td>
<td>2.29 ± 0.37</td>
</tr>
<tr>
<td>20 weeks of exposure + 8 weeks of rest</td>
<td>7.75 ± 0.73*</td>
<td>3.34 ± 0.44*</td>
</tr>
<tr>
<td>+ 20 weeks of exposure</td>
<td>7.49 ± 0.83*</td>
<td>2.32 ± 0.44</td>
</tr>
</tbody>
</table>

| Notation of statistical significance of differences between exposed and control groups as in table 1. |
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Table 4 Some quantitative indices of the state of lungs and TB lymph nodes of exposed and control rats at the end of the 48 week experimental period with different patterns of exposure (X ± Sx)

<table>
<thead>
<tr>
<th>Exposure pattern</th>
<th>Dry weight of lungs (mg/100 g body weight)</th>
<th>Dry weight of TB lymph nodes (mg/100 g body weight)</th>
<th>Lungs Hydroxyproline content (µg/100 g body weight)</th>
<th>Lipid content (mg/100 g body weight)</th>
<th>Dust content (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>No exposure (control)</td>
<td>98.6 ± 4.1</td>
<td>7.1 ± 0.5</td>
<td>1010 ± 36</td>
<td>12.9 ± 0.6</td>
<td>5.7 ± 0.8</td>
</tr>
<tr>
<td>48 weeks of constant exposure</td>
<td>294.7 ± 14.5</td>
<td>39.2 ± 1.2</td>
<td>2426 ± 106</td>
<td>67.2 ± 0.4§</td>
<td>20.8 ± 3.4§</td>
</tr>
<tr>
<td>20 weeks of exposure + 8 weeks of rest + 20 weeks of exposure</td>
<td>218.9 ± 7.2§</td>
<td>29.1 ± 3.0§</td>
<td>1893 ± 73§</td>
<td>48.7 ± 3.1§</td>
<td>16.1 ± 2.1§</td>
</tr>
<tr>
<td>Four subperiods each comprising 10 weeks of exposure + 2 weeks of rest</td>
<td>255.5 ± 10.4§</td>
<td>26.0 ± 1.5§</td>
<td>2114 ± 115§</td>
<td>54.1 ± 4.1§</td>
<td>12.7 ± 1.0§</td>
</tr>
</tbody>
</table>

Notation of statistical significance as in table 1.

between exposures.

The above mentioned mathematical analysis11 can explain this apparent paradox. It was shown that, when total time of exposure subperiods and of rest subperiods are fixed, varying the pattern of their arrangement may decrease the action integral value and increase of the amount of dust retained by lungs only to the moment corresponding to the upper limit of integration—that is, to the moment of estimation of all pulmonary changes. When the AI is taken with limits from zero time to the end of the 48th week, one obtains the following values: 670 mg/wk for the group with one eight week interval in the middle of the period and 690 mg/wk for the group with periodic exposure. Hence, it is just in the first group that one would expect to find a lower intensity of pathological changes and a higher final dust retention.

For one more group used in our experiment—namely, for that given the same eight weeks of rest just before killing (after 40 weeks of uninterrupted exposure)—the computed AI value equals 753 mg/wk. In this group the severity of silicosis differed little from that in the group exposed throughout all 48 weeks and for which the AI is 795 mg/wk, although in the former postexposure elimination of dust had been significant (as was shown above). In other words, the severity was higher than in the two other groups with a total of 40 weeks exposure. (For example, dry weight of lungs and TB lymph nodes in this group were, respectively, 164.3 ± 13.3 and 34.1 ± 2.6 per 100 g body weight—for corresponding data for other groups see table 4. Bearing in mind the long time lag between the last exposure of these rats and their killing, we have not studied BAL in this group as the results of such study would not be comparable with that in the three other groups shown in table 3. Therefore, we did not include results related to this group in table 4.

As to the groups in which the BAL was investigated, the results considered above again justify the conclusion that the observed intergroup differences in the BAL cytology are due not to a dust overload as such but to the severity of the silicotic process induced by it, which in turn corresponds to the AI value. This process takes place mainly in the pulmonary interstices and begins with the phagocytosis of the penetrating silica particles by macrophages of the interstitial pool and with cytotoxic damage to these cells. This pool serves as the main source of AMs recruited to alveoli12 13 and it is logical to suppose that their recruitment may suffer when the damage reaches a certain level. The developing sclerosis of the alveolar septa may also, presumably, interfere with that recruitment.

It is possible to suppose that we deal here not only with inhibited recruitment of AMs but also with a greatly enhanced destruction of recruited AMs by the cytotoxic action of the silica particles phagocytosed on the free surface of the alveoli. Indeed, it was found earlier in our laboratory that during the course of long term inhalation exposure to quartz dust, the proportion of degenerated AMs in the BAL gradually increases.14 Later on we have shown that both the number of NLs in the BAL and the NL/AM ratio increase as the proportion of degenerated AMs increases in silica exposed or control rats, or in dose dependent manner after intratracheal instillation of PMB.1 2 Therefore one may surmise that in the experiment under consideration, a steady rise of both indices reflected the increase in the degeneration of AMs which at the end of the 40th week of exposure had reached such a degree that 24 hours after the last inhalation of dust it was impossible to find the AM count higher than in the controls.

A decrease in the resistance of AMs to the cytotoxic action of silica may, in its turn, be explained if we assume that this cell originated from the already dam-
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Table 5 Main cytological characteristics of bronchoalveolar lavage (BAL) of rats 24 hours after intratracheal instillation of different peritoneal macrophage culture supernatants (X ± Sx)

<table>
<thead>
<tr>
<th>Group of rats</th>
<th>Instilled supernatant after the centrifuging of</th>
<th>No of cells in BAL x 10⁶</th>
<th>Coefficient NL/AM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Total</td>
<td>Alveolar macrophages (AM)</td>
</tr>
<tr>
<td>I</td>
<td>Macrophages</td>
<td>1.02 ± 0.13</td>
<td>0.66 ± 0.12</td>
</tr>
<tr>
<td>II</td>
<td>Macrophages incubated with zymosan</td>
<td>1.65 ± 0.14</td>
<td>0.72 ± 0.17</td>
</tr>
<tr>
<td>III</td>
<td>Macrophages incubated with PMB</td>
<td>1.98 ± 0.30</td>
<td>0.95 ± 0.15</td>
</tr>
<tr>
<td>IV</td>
<td>PMB</td>
<td>1.22 ± 0.12</td>
<td>0.67 ± 0.11</td>
</tr>
<tr>
<td>V</td>
<td>Incubation medium</td>
<td>0.76 ± 0.04</td>
<td>0.52 ± 0.05</td>
</tr>
</tbody>
</table>

*, ‡ and * Values that differ with statistical significance (with p, correspondingly, <0.05, <0.01, and <0.001) from those of group I. § Values of the groups III and V significantly (p < 0.05) different from corresponding values of group IV.

Possibly the inhalation of amosite, a which is less cytotoxic than silica, produces a smaller amount of PMB and, hence, induces a smaller recruitment of NLs. In addition, the small size of an NL probably renders it unable to engulf the fibres even of the length which does not hinder their phagocytosis by AM, and this eventually makes the compensation of NL for the degeneration of AM inadequate. It is also possible that the fibrous shape of asbestos particles mechanically hinders their elimination from fixed lung tissues and thus leads to the phenomenon of “sequestration” which was not observed after exposure to quartz dust. These explanations of the kinetic differences between amosite and quartz are, however, no more than speculative.

A cause effect relation between the enhancement of the degeneration of AMs in the lower airways and that of the recruitment of NLs we explained formerly by a direct NL attractant effect of PMB. This effect has been repeatedly shown in experiments with intratracheal injection of whole PMB or PMB supernatant and with resuspended lipids extracted from PMB and also in experiments in vitro which showed the stimulation of the mobility of both NLs and AMs in the presence of PMB. Later, experimental results of American investigators became available which showed that activated alveolar macrophages are able to produce, or to release, a chemotactic factor for human blood neutrophils. We had accumulated many experimental data suggesting that PMB activate a viable macrophage and we supposed that the preferential recruitment of NLs to the airways might be due not only to the above mentioned attractant action of PMB themselves but also to the PMB influencing viable AMs so as to make them produce or release a specific NL attractant.

As seen from table 5, the activation of peritoneal macrophages with zymosan in vitro carried out under the same condition as in the experiments of the American authors resulted in a significant increase in the ability of the cellular culture’s supernatant instilled intratracheally to recruit NLs into the airways, but this activation did not influence the aged interstitial pool. It would be impossible to deny the possibility, however, that changes in cell resistance also reflect some general shift in the host’s non-specific reactivity. It has been shown in our laboratory that on the one hand, the resistance of the AM to the cytotoxicity of silica in vivo could be predictably moderated by agents and factors influencing the host’s general reactivity, and on the other, that the latter underwent biphasic shifts on the background of the development of silicosis may, in turn, be decreasing when this development reached a sufficiently advanced stage.

The enhanced recruitment of NLs may, in turn, be due not only to the intensified breakdown of AM— that is, to the increase of endogenous PMB dose—but also to some changes in the host’s ability to react to the PMB stimulus. For example, we showed earlier that the recruitment of NLs after intratracheal instillation of a fixed PMB dose increased against the background of the influence of a mineralocorticoid, or of cholinomimetic or adrenoblocking effects. In other words, one may suppose that the compensatory increase in recruitment of NLs observed in the present investigation was mediated not only through self regulation but through neurohormonal regulatory mechanisms as well.

Having analysed retrospectively some earlier experimental data in the light of the present results, we found that they are in good agreement with the conjectures discussed above. For instance, in rats inhaling the same quartz dust although in a lower concentration (47.9 ± 2.9 mg/m³) the NL/AM ratio was 2.94 times as great as in control rats after 10 weeks, and 10.00 times after 33 weeks exposure. Still more pronounced was the decrease in a group exposed to the far more cytotoxic aerosol of colloidal silicic acid (52.8 ± 4.1 mg/m³): respectively, 7.06 and 31.50 times. In the second group pulmonary silicon dioxide retention has not increased during the course of the investigation; it had reached a “plateau” level already by the 10th week of exposure, which fact corroborates our hypothesis of a constant clearance rate maintained by the increased contribution from the NLs.
Some peculiarities of the pulmonary phagocytic response

Some peculiarities of the pulmonary phagocytic response. One may conclude that the ability of an activated cell to produce an NL attractant is characteristic not only of the alveolar but also of the peritoneal macrophage, and that this ability may be shown not only with in vitro experiments modelling chemotaxis21–24 but also with intrathecal instillation of macrophageal supernatant.

The supernatant of PMB, as in our earlier experiments, induced the recruitment of phagocytic cells into airways with an increased NL/AM ratio (table 5). Greater shifts in the same direction were induced with the supernatant of viable macrophages preincubated with the same PMB dose, this response being also significantly stronger than that of the supernatant of macrophages preincubated without PMB. The cellularity of the BAL and the NL/AM ratio after instillation of the latter did not differ significantly from those after instillation of a control incubation medium. (Resting macrophages do not produce an NL attractant and their breakdown during pre-incubation and centrifuging was evidently not sufficient to give the same effect as PMB.)

Taken as a whole, the results of this experiment seem to be in good agreement with our hypothesis of the production or release of an NL attractant by PMB activated viable macrophages.

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Some peculiarities of the pulmonary phagocytotic response: dust retention kinetics and silicosis development during long term exposure of rats to high quartz dust levels.

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*Br J Ind Med* 1987 44: 228-235
doi: 10.1136/oem.44.4.228

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