Influence of local vibration on plasma creatine phosphokinase (CPK) activity

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ABSTRACT This study was designed to obtain basic information about the mechanism of the occurrence of muscular disorders after exposure to vibration. The hind legs of rats were exposed to acute and chronic local vibration at frequencies of 30, 60, 120, 240, 480, and 960 Hz with a constant acceleration of 50 m/sec². The exposure time was four hours for acute, and four hours a day for two weeks continuously for chronic exposure. Blood was collected after exposure to measure plasma creatine phosphokinase (CPK) activity. In both exposure groups the activity of plasma CPK was significantly higher at 30, 60, 120, 240, and 480 Hz compared with the control group and was especially high at 30 Hz; there was no significant change at 960 Hz. As a result of an analysis of the CPK isoenzymes, the increase in plasma CPK activity was shown to be due to the activity of the plasma CPK-MM fraction, originating in the skeletal muscle. Plasma CPK activity showed a tendency to decrease gradually with the increase in vibration frequency during acute exposure but showed no such tendency during chronic exposure. There was no remarkable pathohistological change in muscle preparations from the hind legs, hence it was presumed that the increase in plasma CPK activity was caused not by the morphological changes of muscle but by other mechanisms, such as an increase in the permeability of the cell membrane.

The hazards to operators using vibrating tools such as chain saws or rock drills are caused by hand-arm transmitted vibration. The typical symptoms are based on disorders of the peripheral circulation, the peripheral nerves and the musculoskeletal system. Concerning the disorders of muscle, it is controversial whether the disorders arise from vibration itself or from other causes related to the working conditions. Although some epidemiological studies relating to this problem have been reported, there have been few experimental studies. Considering vibration as a physical stress, it is important to take account of the intensity and frequency of vibration. The vibrations from vibrating tools are composed of different frequencies so that it seems useful to indicate which frequency has a serious effect on muscle function in order to help elucidate the mechanism by which the disorders are caused.

The purpose of the present study was to obtain some basic information about the mechanism of the occurrence of muscle disorders that are caused by exposure to local vibration.

Material and methods

MATERIALS
For acute local vibration exposure experiments, the animals used were 35 male Wistar strain rats weighing 246–302 g; for chronic vibration exposure experiments, 33 male rats of the same strain were used.

EXPOSURE TO VIBRATION
The apparatus used to produce vibration was composed of an electromagnetic shaker (EMIC 513-A, shaking power 7·5 kg; vibration frequency range 5–5000 Hz) coupled to an amplifier (TACHIKAWA TA-100), a function oscillator (TORIO AG202), and a vibration meter (EMIC 505-D).

The animals were placed in individual wire mesh cages, in the prone position. The hind legs were outside the case and the plantar surfaces were horizontally fixed to the vibrating plate by means of double sided adhesive tapes so that the vibration would be transmitted only to the hind legs. The part of the cage
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containing the rest of the rat was fixed on a non-vibrating plate separated from the shaker. The hind limbs were exposed to vertical sinusoidal vibration with frequencies of 30, 60, 120, 240, 480, and 960 Hz under a constant acceleration of 50 m/sec^2. The duration of exposure was four hours in the acute experiments, and four hours a day for two weeks in the chronic exposure experiments.

Before the experiments the rats were trained for one week to ride on the vibrating platform.

The control group for each experiment was placed in wire mesh cages like the exposed group and the cages were placed near to the operating electromagnetic shaker during the exposure experiment, so that any noise effect would still be experienced.

**BLOOD COLLECTION**

Shortly after the exposure in the acute experiment, and 24 hours after the last exposure in the chronic experiment, pentobarbital (0.8 mg/kg weight) was intraperitoneally injected into the rats and arterial blood was drawn from the abdominal aorta by means of heparinised syringe under anaesthesia. The collected blood was centrifuged for 20 minutes at 2000 rpm and the plasma was separated.

**ASSAY OF PLASMA CREATINE PHOSPHOKINASE (CPK) ACTIVITY**

The plasma total-CPK(t-CPK) and its isoenzyme, plasma CPK-MM fraction, were separated using a CPK assay kit (Worthington Biochemical Corporation, USA) and their activities were determined using a spectrophotometer (Shimazu UV 202 type). Each enzyme activity was determined within 12 hours of collecting the blood samples.

**PATHOHISTOLOGICAL OBSERVATION**

Pathohistological preparations of muscles from the hind legs were made from the chronically exposed groups (30, 60, and 240 Hz) and the controls, stained with hematoxylin and eosin, and examined with a light microscope.

**Results**

**ACUTE LOCAL VIBRATION EXPOSURE**

Figure 1 shows the values of plasma t-CPK activity at each vibration frequency, expressed as the mean ± SD. The values for the plasma t-CPK activity in the exposed groups were 482.4 ± 62.4 IU/l at 30 Hz, 249.0 ± 82.5 IU/l at 60 Hz, 232.4 ± 53.8 IU/l at 120 Hz, and 141.9 ± 28.9 IU/l at 240 Hz. Compared with the value in the control group (33.1 ± 14.2 IU/l) each of those in the exposed groups was significantly high (p < 0.001). The value at 480 Hz (86.9 ± 22.4 IU/l) was also shown as high by comparison with the control (p < 0.01). The value at 960 Hz was 51.2 ± 16.4 IU/l and was not significantly different from the control value.

Figure 2 shows the values for the plasma CPK-MM fraction activity at each vibration frequency; the figures are expressed as the mean ± SD. The values
were 467 ± 50·2 IU/l at 30 Hz, 243·1 ± 81·2 IU/l at 60 Hz, 228·5 ± 53·9 IU/l at 120 Hz, and 136·4 ± 29·3 IU/l at 240 Hz. Compared with the control value (31·6 ± 13·6 IU/l) each of the exposed levels was significantly high (p < 0·001). The value at 960 Hz was not significantly different from the control value.

**CHRONIC LOCAL VIBRATION EXPOSURE**

Figure 3 shows the values of plasma t-CPK activity for each vibration frequency, which are expressed as the mean ± SD. The plasma t-CPK activities of the exposed group were 430·0 ± 41·9 IU/l at a vibration frequency of 30 Hz, 204·6 ± 35·2 IU/l at 60 Hz, 181·0 ± 38·4 IU/l at 120 Hz, 186·1 ± 38·4 IU/l at 240 Hz, and 249·5 ± 82·5 IU/l at 480 Hz. Compared with the value of the control group (93·8 ± 8·5 IU/l) each of these was significantly high (p < 0·001). The plasma t-CPK activity at 960 Hz (106·7 ± 34·4 IU/l) was not significantly different from the control, however.

![Figure 3](image-url)  
*Fig 3 Effects of different vibration frequencies on plasma t-CPK activity (mean ± SD). Figures in parentheses indicate number of rats. *p < 0·001.*

Figure 4 shows the values of the activity of the plasma CPK-MM fraction for each vibration frequency (mean ± SD). The values were 422·4 ± 42·5 IU/l at 30 Hz, 198·2 ± 35·2 IU/l at 60 Hz, 175·2 ± 29·4 IU/l at 120 Hz, 178·8 ± 30·0 IU/l at 240 Hz, and 240·8 ± 76·4 IU/l at 480 Hz. Each value was significantly higher than that of the control group (86·5 ± 7·8 IU/l) (p < 0·001). The CPK-MM fraction activity at 960 Hz was 98·9 ± 34·1 IU/l and this was not significantly different from the control value.

![Figure 4](image-url)  
*Fig 4 Effects of different vibration frequencies on plasma CPK-MM fraction activity (mean ± SD). Figures in parentheses indicate number of rats. *p < 0·001.*

Concerned with the microscopic examination of the preparations from the hind legs, no pronounced pathohistological difference was observed between the control and the group chronically exposed to vibration. The pathohistological pictures of the plantaris muscle exposed (30 Hz) and the control groups are shown in fig 5.

**Discussion**

There have been many reports3–7 of the disorders of muscle, bone, or joint systems caused by using vibration tools since Holzman's original observations in 1929.8 Although some general research on these muscle disorders has been undertaken,7 there is little information as to the mechanism by which they are caused.

The present study was designed to investigate the influence of local vibration on muscle function. We focused on CPK which is abundant in the skeletal muscle, and tried to clarify whether or not the activity of this enzyme is affected by vibration and to estimate which vibration frequency is most effective at inducing muscle disorders. The plasma t-CPK activity showed significantly high values at 30, 60, 120, 240, and 480 Hz compared with the control with both acute and chronic exposure to local vibration. The increase in plasma t-CPK activity was found to be due to that of the plasma CPK-MM fraction which originated in the skeletal muscle. At frequencies of 30, 60,
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120, 240, and 480 Hz, a definite increase in plasma CPK activity was observed with both acute and chronic exposure. The highest activity was found at 30 Hz with both types of exposure. Another point of interest was that, with acute exposure, the increase in plasma t-CPK activity showed a tendency gradually to decrease with the increase of vibration frequency. With chronic exposure, no such tendency was observed. At 480 Hz the responses of acute exposure and chronic exposure were remarkably different. Thus we were interested to consider the effects of local vibration on plasma CPK activity.

The activity of sarcoplasmic enzymes is increased in various sarcoplasmic diseases such as progressive muscular dystrophy and polymyositis, and in other diseases including myocardial infarction. Some reports have also commented on the changes in sarcoplasmic enzymes in relation to exercise or heat stress. By contrast, there have been few reports concerned with the influence of vibration on sarcoplasmic enzymes. It is of considerable practical significance that an increase in CPK activity was found to be induced by vibration and that characteristic changes in CPK activity were noted with different vibration in the present experiment.

Two possibilities may be considered with respect to the increase in CPK activity; the effusion of CPK from the sarcoplasm into the plasma caused by muscular tissue destruction and a rise in the permeability of the muscle cell membrane due to local vibration. In the present experiment pathohistological investigation of the hind legs of rats subjected to the vibration showed no structural changes suggesting muscular tissue destruction. Therefore, presumably, the increase in plasma CPK activity induced by local vibration is attributable to a rise in the permeability of the muscle cell membrane.

References

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